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Full Length Research Paper

# The effects of zinc (Zn) and C<sup>14</sup>-indoleacetic acid (IAA) on leaf senescence in *Helianthus annuus* L.

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Sequential leaf senescence is defined as a kind of programmed death events which is an important process in growth of plant. This study aimed to explore the sequential leaf senescence rate due to indoleacetic acid and lack of zinc (-Zn). Therefore, the effect of zinc and indole-3-acetic acid on senescence which occurs in *Helianthus annuus* (sunflower) cotyledons was analyzed. It was found that in cotyledons of seedlings grown in Hoagland solution which was prepared without addition of zinc senescence is delayed. It was recognised that in case of <sup>14</sup>C indoleacetic acid (IAA) which was given from apical tip not reaching the root and cotyledons, senescence does not occur in cotyledons. It was studied to get more information about physiological system of sequential leaf senescence.

**Key words:** Sequential leaf senescence, cotyledon, zinc, <sup>14</sup>C indole-acetic acid (IAA), *Helianthus annuus* (sunflower).

### INTRODUCTION

Senescence is the final phase of plant vegetative and reproductive development, preceding the widespread death of cells and organs (Schmid et al., 1999; Guiboileau et al., 2010; Caswell and Salguero-Gomez, 2013). It has long been known that hormones regulate the progression of leaf senescence (Fletcher and Osborne, 1965; Misra and Biswal, 1980; Noodén and Leopold, 1988; Jibran et al., 2013). In the process of senescence, destruction cases occur more than synthesis. From point of that view, definition of senescence is the process which increases destruction cases in cell and causes the plant to die.

The analysis made on leaf cells shows that during senescence consecutive metabolic events occur. These

events can be ordered as the synthesis of proteolitic enzyme (Colin and Thimann, 1972; Cheng and Kao, 1984; Hörtensteiner and Feller, 2002), the start of destruction of membrane proteins caused by these enzyme's activities, the decrease of quantity of protein and total nitrogen in the cell (Krul, 1974; Peterson and Huffaker, 1975; Peoples and Dalling, 1978, Prakash et al., 2001; Hopkins et al., 2007; Kaplan-Dalyan and Sağlam-Çağ, 2013), the acceleration of chlorophyll destruction (Peoples et al., 1980; Rodoni et al., 1997; Hörtensteiner, 2006; Darnel et al., 1990) and lipid destruction (Dhindsa et al., 1982; Harwood et al., 1982; Thompson et al., 1998; Hebeler et al., 2008). It is accepted that transportation of nutrients in other leaves

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starting from the oldest organ to the youngest supports the nutrient drain hypothesis. According to another hypothesis called 'signal' hypothesis, a signal which is thought to be synthesized by developing seeds, is being transported to old leaves and causes senescence as a result of catabolic reactions. According to this hypothesis, if the signal center is eliminated, senescence does not occur (Lindoo and Noodén, 1977). But the above mentioned signal has been displayed that it could not be isolated (Moore, 1979; Ridge, 1991). It is obviously known that the cause of all biochemical events during senescence are releated to gene expression (Draper, 1969; Sanders and Write, 1995; Hörtensteiner, 1997; Distelfeld et al., 2014). The meanings of these chemical events come into being only with the researches made by plant physiologs on plant's physiology. While it is being said that auxins (Wareing and Seth, 1967; Kahanak et al., 1978; Lim et al. 2003) delay senescence, researches made recently indicate that auxins (Palni et al., 1988; Lu et al., 2001; Gören and Sağlam-Çağ, 2007) accelerate the senescence. Otherwise, Noh and Amasino (Noh and Amasino, 1999) detected that auxin represses transcription of some genes whose expression is correlated with senescence.

### **MATERIALS AND METHODS**

Helianthus annuus L. seedlings were grown in intensity of 6000 lux light, under 12 h photoperiod and  $26 \pm 2^{\circ}$ C.

### Designation of senescence degree

To determine the senescence which occur in cotyledons of *H. annuus* quantitatively the method improved and used for soya bean's Anoca variety-show by Lindoo and Noodén (1976) was adapted to *H. annuus* cotyledons. For chlorophyll designation Arnon (1949) method was used. To determine total nitrogen quantity a method, formed with combination of Kjeldahl method and spectrophometric measurement method was used (Lindoo and Noodén, 1976). The zinc quantity in the material was designated with atomic absorption spectrophotometer (AAS).

### Giving IAA to the cut ends of the decapitated seedlings

One to two days before senescence starts in cotyledons, seedlings were decapitated by being cut approximately 3 cm above the internodium cotyledons.  $10^{-5}$  M. IAA solution (treated) or water (control) was applied to decapitated surface.

### Giving <sup>14</sup>C-IAA to the seedlings on the top buds and nodium leaves

 $10^{-5}$  mol.  $^{14}$ C-IAA (specific activity:40 mCi / mmol.) 1 drop 1% tween-20 was added per 1 ml. 60 and 120 µl from IAA solution was dropped on plant's top bud, 80 and 320 □l was dropped on nodium leaves. To hinder indolacedic acid's photo oxidation, this process was made when the plants were passing to dark period. The whole organ, of which radioactivity will be enumerated in β counter was

prepared accordingly. Counting value of the material per 5 min (cp5m=count per 5 min) was calculated.

#### Statistic evaluation of the results

Standard deviation estimate was made to evaluate the results obtained from experiment and control groups statistically. In case of the number obtained when  $\pm$  values of differences between the result's square's total sum's square root is multiplied with three is found to be smaller than the difference between values, the difference is decided to be significant statistically.

### **RESULTS**

As it is known that zinc provides indole-acetic acid (IAA) stabilization and in case of zinc deficiency quantity of IAA decreases, this mineral's effects on plant growing and cotyledon senescence were analysed. The seedlings forming the experiment group were grown in Hoagland solution which does not include zinc and was diluted in 1/8 ratio (Table 1). Senescence delayed in cotyledons of seedlings in -Zn solution. Besides with the ingathering, when cotyledon senescence in plants grown with the existence of zinc (control) is 50% according to plastochron index (28th day), total chlorophyll and total nitrogen quantities in cotyledons of all seedlings belonging to experiment or control group (Table 2). The quantity of the zinc which is thought to exist in the seed naturally was measured with AAS (Figure 1). Nineteen days old H. annuus seedlings were devided into 4 groups. First group plants were intact (control). Other group plants were given IAA, NAA and H<sub>2</sub>O from truncated end being decapitated from under 2nd internodium. After this process the speed of senescence occuring in cotyledons was observed (Figure 2). In cotyledons of plants to which IAA and NAA applied senescence occured quickly just like it occurs in cotyledons of intact plants. However, in a great majority of plants having a process with H<sub>2</sub>O, cotyledons remained green. Senescence did not occur in the cotyledons of the 17 days old experiment and control seedlings which were exposed to the same process and application. To determine first which organ as a target indolacedic acid after being produced in the plant is transported, 120 µl from 10<sup>-5</sup>M <sup>14</sup>C-IAA+tween 20 upon top bud of the plants was dropped in. After dark period for 12 h, the quantity of <sup>14</sup>C-IAA in the roots and cotyledons of the plants was stated (Table 3). Radioactivity existing in the root is found to be more than 20 times more than the radioactivity in cotyledons. From the values obtained it was understood that IAA given from top bud was transported quickly to the root. On the other hand, from the middle of the first internodium's of the seedlings, a part, approximately 1 cm was boiled with hot water vapour on the 17 day 60  $\mu$ l  $10^{-5}$  M-IAA+tween 20 was dropped in the top bud of the plants of which cotyledons was just 100% gren on the 32 day. After dark period for

Day	Average green area Percentage [Control (+ Zn)]	Day	Average green area Percentage [Control (- Zn)]
17	100.00 ± 0.00	20	100.00 ± 0.00
22	$78.00 \pm 0.26$	23	73.93 ± 1.62
25	56.34 ± 1.42	27	49.45 ± 1.23
29	$32.73 \pm 2.46$	30	21.61 ± 1.72
33	$00.00 \pm 0.00$	34	$00.00 \pm 0.00$

**Table 1.** The effect of Zn (105 mg/L) on the green area (%) of the cotyledons from-day *H. annuus* seedlings grown in 1/8 Hoagland solution.

**Table 2.** The effect of Zn (105 mg/L) on the chlorophyll and nitrogen content of the cotyledons from 28 day *H. annuus* seedlings grown in 1/8 Hoagland solution.

Hoagland	mg N / g cotyledon	%	mg chlorophyll / cotyledon	%
Hoagland (- Zn)	$129.603 \pm 4.256$	100	$0.0449 \pm 0.005$	100
Hoagland (+ Zn)	$108.109 \pm 2.562$	83.4	$0.0382 \pm 0.002$	74.2

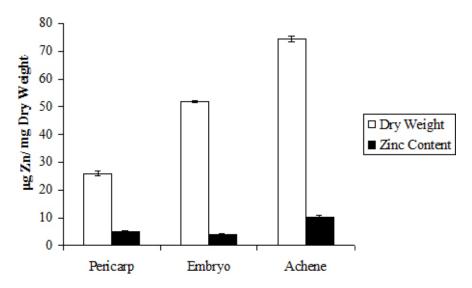


Figure 1. Dry weight and Zn quantity in pericarp, embryo and achene.

12 h, radioactivity in different organs of the plants was measured (Table 4).

As the radioactivity difference counted in organs under the 1 internodium's boiled part was unsignificant statistically, it was understood that radioactivity can pass downwards from boiled area in trace quantity. Besides, althought it was found on boiled part, on the leaves of the 2 internodium a statistically significant quantity of radioactivity could not be found. It was stated that radioactivity had accumulated in a great quantity on the boiled part of internodium.

To the first leaves (second nodium leaf) after cotyledon of 19 days old plants totally  $80\mu l^{14}$ C-IAA, 40 per each, was applied. After plants being ingathered on different

days (Avery et al., 1937; Lindoo and Noodén, 1976; Papadopoulus et al., 1985; Noodén and Leopold, 1988), radioactivity on root and 3. nodium leaves was measured (Table 5).

It was recognised that <sup>14</sup>C-IAA applied to 2 nodium leaves was transported to root first and then from there by xylem, was transported to the leaves making transpiration.

### DISCUSSION

In this research, the effects of zinc and indolacedic acid, which is a growing hormone, on senescence was

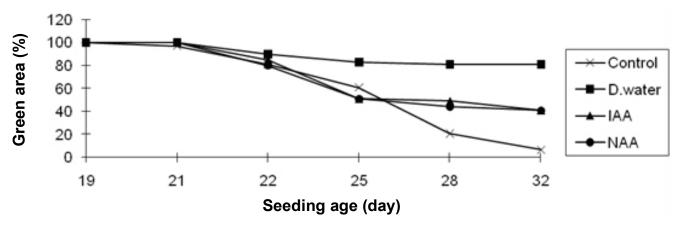


Figure 2. Green area (%) of cotyledons of 19 days old H. annuus seedlings which were decapitated below the second internode and treated with  $10^{-5}$ M IAA,  $10^{-5}$ M NAA or  $H_2$ O from the truncated end.

**Table 3.** <sup>14</sup>C amounts in the roots and cotyledons of *H. annuus* seedlings treated with 10<sup>-5</sup> M <sup>14</sup>C-IAA (\* Significant).

Organ	Count / 5 min	Radiation of background	Difference
Cotyledon	299.85 ± 31.25	$288.36 \pm 3.015$	11.49
Root	875.41 ± 29.09	$288.36 \pm 3.015$	587.05 *

Table 4. <sup>14</sup>C amounts in different organs of *H. annuus* seedlings treated with 10<sup>-5</sup> M <sup>14</sup>C-IAA (\* Significant) .

Organ	Count / 5 min	Radiation of background	Difference
2nd node leaves	$306.92 \pm 7.05$	$288.36 \pm 3.015$	18.56
First internode	$1039.33 \pm 57.63$	$288.36 \pm 3.015$	750.97 *
100% Green cotyledon	$299.87 \pm 7.66$	$288.36 \pm 3.015$	11.51
Hypocotyl	$304.00 \pm 11.82$	$288.36 \pm 3.015$	15.64
Root	$313.00 \pm 10.64$	$288.36 \pm 3.015$	24.64

**Table 5.** <sup>14</sup>C amounts in the roots and 3<sup>rd</sup> node leaf of *H. annuus* seedlings leaf on different days after 10<sup>-5</sup> M <sup>14</sup>C-IAA treatments on the 2<sup>nd</sup> nodium. (\* Significant).

Day	Root	3rd nodium leaf	Radiation of	Difference	Difference
	(Count / 5 min)	(count / 5 min)	background	(root)	(3 rd nodium leaf)
20	$318,44 \pm 11.627$	$302,67 \pm 3.567$	$288.36 \pm 3.015$	30.08	14.31 *
21	$489,11 \pm 34.310$	$352,22 \pm 10.97$	$288.36 \pm 3.015$	200.75*	63.86 *
22	$454.00 \pm 45.287$	$342.00 \pm 14.09$	$288.36 \pm 3.015$	165.64 *	53.64 *
25	485,67 ±11.794	$712,89 \pm 131.27$	$288.36 \pm 3.015$	197.31 *	424.53 *

searched. In a reseach (Ray and Choudhuri, 1981), it was supported that hormones (IAA, GA, Kinetin) plays the most important role in transporting nutrients to seeds that develops as an endogenic hormone resource. It is known that the deficiency of the hormones which perevents senescence (for ex: cytokinin) may cause senescence. Palni et al. (1988) mentions that auxin has

an effect on cytokinin's metabolism and this effect is actualized by oxidase enzyme. While some researchers (Jacobs and Cready, 1967; Sanchez-Bravo et al., 1991) declare that indolacedic acid localize in cortex, vascular tissue and pith, auxin is transported in vascular and epidermal tissues, other researchers (Bangerth, 1994; Ekölf et al., 1995; Li et al., 1995; Shimizu-Sato et al., 2009)

emphasized that intact plants's cytokinins in xylem exudate are under the control of polar auxin transportation system. Hare and Staden (1994) expressed that cytokinin catabolism which becomes true with the activity of sitokinin, a specific enzyme oksidase realizes death in plant tissue and moreover stated that auxin plays the role of allosteric systematizer increasing this enzyme's activity.

It is known that indolacedic acid is synthesized in the end of stem and zinc provides the stabilization of indolacedic acid (Skoog, 1940; Takaki and Kushizaki, 1970). In the early development phase of the plants of which endogen IAA quantity was decreased by being grown in zinc deficiency, IAA that is under the control of the quantity of zinc in the seed has such a quantity that it delays the senescence but can provide growing. But zinc which is given with Hoagland solution in addition to the zinc quantity in the seedling may be impulsive in senescence or may delay growing because of its toxical effects on some enzyme systems releated to growing. Likewise, Sağlam-Çağ and others (Sağlam-Çağ et al., 2004) emphasized that senescence was delayed in excised cotyledons in the solution lacking zinc. In that research, in the existence or deficiency of zinc, IAA which can be controlled endogenly was hold to be responsible for the change in senescence's speed. In some experiments which <sup>14</sup>C-IAA was used (Hew et al., 1967), it was noticed that IAA given from truncated end of the stem goes through stem axis quickly and don't enter to leaves. Also, in this research it was found that <sup>14</sup>C-IAA was transported to root without touching at leaves. Moreover, as <sup>14</sup>C's internodium does not goes through boiled part, it was noticed that it could not reach the root and cotyledons and senescence didn't occur in cotyledons.

We can assert that sequential leaf senescence is releated to the occurance of metaxylem after protoxylem and in this event, with IAA's effect on xylem formation, it may come on the scene. Just before senescence, although senescence occured when IAA was applied from truncated top, cotyledons remained green when IAA was applied in early phase. Researhers (Shimomura et al., 1988; Jones et al., 1989; Jones, 1994) indicated that there are 2 different receptor in plasma membrane connecting IAA and in recent years it was determined that first one of these receptors isolated is releated to cell growing but then any absolute information about second IAA receptor's function wasn't given (Darnel et al., 1990; Cooper, 1997).

As a result of our research, we saw that zinc, providing IAA stabilization accelerates senescence; in the researches made with <sup>14</sup>C, as its internodium does not go through boiled part <sup>14</sup>C-IAA given from apex, it can not reach root and cotyledons and senescence does not occur in cotyledons. It became certain that it was transported to the root without touching at leaves and this transportation is made by parenchymatic living tissues not xylem. This research indicated that senescence signal may be indoleacetic acid or a substance like indoleacetic acid.

#### Conflict of interests

The author(s) have not declared any conflict of interests.

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### **REFERENCES**

- Arnon DI (1949). Copper enzymes in isolated chloroplast. Polyphenoloxidase in *Beta vulgaris*. Plant Physiol. 24:1-15.
- Avery GS, Burkholder Jr. PR, Creighton HB (1937). Nutrient deficiencies and growth hormone concentration in *Helianthus* and *Nicotiana*. Am. J. Bot. 24:553-57.
- Bangerth F (1994). Response of cytokinin concentration in the xylem exudate of bean (*Phaseolus vulgaris* L.) plants to decapitation and auxin treatment, and relationship to apical dominance. Planta 194:439-442.
- Caswell H, Salguero-Gomez R (2013). Special feature new perspectives in whole-plant senescence: Age, stage and senescence in plants. J. Ecol. 101:585-595.
- Cheng SH, Kao CH (1984). The role of proteolytic enzymes in protein degradation during senescence of rice leaves. Physiol. Plantarum 62:231-237.
- Colin M, Thimann K (1972). Role of protein synthesis in the senescence of leaves. Plant Physiol. 50:432-437.
- Cooper GM (1997). The cell: A molecular approach. ASM Press. Washington, D.C. Sinauer Associates, Inc.Sunderland, Massachusetts. pp. 521-560.
- Darnel J, Lodish H, Baltimore D (1990). Molecular Cell Biology. Second Ed. Scientific American Books. W.H. Freeman Company, Newyork. p. 709-762
- Dhindsa RS, Plumb-Dhindsa PL, Reid DM (1982). Leaf senescence and lipid peroxidation: Effects of some phytohormones, and scavengers of free radicals and singlet oxygen. Faculdade de Agronomia, Univ. Eduardo Mondlane, Maputo, Mozambique, Department of Biology, University of Calgary, Calgary, Alberta T2N 1N4, Canada.
- Distelfeld A, Avni R, Fischer AM (2014). Senescence, nutrient remobilization, and yield in wheat and barley. Jour. Experiment. Bot. 27:1-16.
- Draper SR (1969). Lipid Changes in Senescing Cucumber Cotyledons. Phytochemistry 8:1641-1647.
- Ekölf S, Astot C, Blackwell J, Moritz T, Olsson O, Sandberg G (1995). Auxin/cytokinin interactions in wild-type and transgenic tobacco. Plant Cell Physiol. 38:225-235.
- Fletcher RA, Osborne DJ (1965). Regulation of protein and nucleic acid synthesis by gibberellin during leaf senescence. Nature 207:1176-
- Gören N, Sağlam-Çağ S (2007). The Effect of Indole-3-acetic acid and benzyladenine on sequential leaf senescence on *Helianthus annuus* L. seedlings. Biotechnol. Biotec. Eq. 21:322-327.
- Guiboileau Å, Sormani R, Meyer C, Masclaux-Daubresse C (2010). Senescence and death of plant organs: nutrient recycling and developmental regulation. CR. Biol. 333:382-91.
- Hare PD, Van Staden J (1994). Cytokinin Oxidase: Biochemical features and physiological significanse. Physiol. Plant., 91:128-136.
- Harwood JL, Jones AVHM, Thomas H (1982). Leaf Senescence in a non-yellowing mutant of *Festuca pratensis*. III. Total acyllipids of leaf tissue during senescence. Planta. 156:152-157.
- Hebeler R, Oeljeaklaus S, Reidegeld KA, Eisenacher M, Stephan C, Warscheid B (2008). Study of early leaf senescence in *Arabidopsis thaliana* by quantitative proteomics using reciprocal 14N/15N labeling and difference gel electrophoresis. Mol. Cell Proteomics 7:108-20.
- Hew CS, Nelson CD, Krotkow G (1967). Hormonal kontrol of translocation photosynthetically assimilated <sup>14</sup>C in young soybean plants. Am. J. Bot. 54:252-256.

- Hopkins M, Taylor C, Liu Z, Ma F, McNamara L, Wang T, Thompson JE (2007). Regulation and execution of molecular disassembly and catabolism during senescence. New Phytol. 175:201-214.
- Hörtensteiner S (1997). Chlorophyll breakdown in senescent chloroplasts (cleavage of pheophorbide a in two enzymic steps). Plant Physiol. 115:669-676.
- Hörtensteiner S (2006). Chlorophyll degradation during senescence. Annu. Rev. Plant Biol. 57:55-77.
- Hörtensteiner S, Feller U (2002). Nitrogen metabolism and remobilization during senescence. J. Exp. Bot. 53:927-937.
- Jacobs WP, Mc Cready CC (1967). Polar transport of growth-regulators in pith and vascular tissues of coleus stems. Am. J. Bot. 54:1035-1040.
- Jibran R, Hunter DA, Dijkwel PP (2013). Hormonal regulation of leaf senescence through integration of developmental and stress signals. Plant Mol. Biol. 82:547-561.
- Jones AM (1994). Auxin-binding Proteins. Annu. Rev. Plant Phys. 45:393-420.
- Jones AM, Lamerson P, Venis MA (1989). Comparison of Site I Auxinbinding and a 22-kilodalton Auxin-binding Protein Maize. Planta 179:409-413.
- Kahanak GM, Okatan Y, Rupp DC, Noodén LD (1978). Hormonal and genetic alteration of monocarpic senescence in soybeans. Plant Physiol. 61:26.
- Kaplan-Dalyan E, Sağlam-Çağ S (2013). The effect of epibrassinolide on senescence in horizontal sunflower (*Helianthus annuus* L.) seedlings. IUFS J Biol, 72(1):33-44
- Krul WR (1974). Nucleic acid and protein metabolism of senescing and regenerating soybean cotyledons. Plant Physiol. 54:36-40.
- Li CJ, Guevara E, Herrera J, Bangerth F (1995). Effect of apex excision and replacement by 1-naphthylacetic acid on cytokinin concentration and apical dominance in pea plants. Physiol. Plantarum 94:465-469.
- Lim PO, Woo HR, Nam HG (2003). Molecular genetics of leaf senescence in Arabidopsis. Trends Plant Sci. 8:272-278.
- Lindoo SJ, Nooden LD (1977). Studies on the behavior of the senescence signal in *anoka soybeans*. Plant Physiol. 59:1136-1140.
- Lindoo SS, Noodén LD (1976). The interrelation of fruit development and leaf senescence in anoka soybeans. Bot. Gaz. 137:218-223.
- Lu IL, Sutter E, Burger D (2001). Relationships between benzyladenine uptake, endogenous free IAA levels and peroxidase activities during upright shoot induction of Cymbidium ensifoilum cv. Yuh Hwa rhizomes in vitro. Plant Growth Regul. 35:161-70.
- Misra AN, Biswal UC (1980). Effect of phytohormones on chlorophyll degradation during aging of chloroplasts in vivo and in vitro. Protoplasma 105(1-2):1-8.
- Moore TC (1979). Biochemistry and physiology of plant hormones. In chapter 6 Ethylene. Springer Verlag, New York. 208-229.
- Noh YS, Amasino RM (1999). Identification of a promoter region responsible for the senescence-specific expression of SAG12. Plant Mol. Biol. 41:181-194.
- Noodén LD, Leopold AC (1988). Senescence and aging in plants. Academic press, San Diego.
- Palni LMS, Burch L, Horgan R (1988). The effect of auxin concentration on cytokinin stability and metabolism. Planta 174:231-234.
- Papadopoulus I, Rending VV, Broadbent FE (1985). Growth, nutrition and water uptake of tomato plants with divided roots growing in differentially salinized soil. Agron. J. 77:21-26.

- Peoples MB, Beilharz WC, Waters SP, Simpson RJ, Dalling MJ (1980). Nitrogen redistribution during grain growth in wheat (*Triticum aestivum* L.). II. Choloroplast senescence and the degradation of ribulose-1,5-BiP carboxilase. Planta 149:241-251.
- Peoples MB, Dalling MJ (1978). Degradation of ribulose-1,5 biphosphate carboxylase by proteolytic enzymes from crude extract of wheat leaves. Planta 138:153-160.
- Peterson LW, Huffaker RC (1975). Loss of ribulose 1,5 diphosphate carboxilase and increase in proteolytic activity during senescence of detached primary barley leaves. Plant Physiol. 55:1009-1015.
- Prakash JSS, Baig MA, Mohanty P (2001). Senescence induced structural reorganization of thylakoid membranes in Cucumis sativus cotyledons; LHC II involvement in reorganization of thylakoid membranes. Photosynth. Res. 68:153-161.
- Ray S, Choudhuri MA (1981). Effects of Plant Growth Regulators on Grain-filling and Yield of Rice. Ann. Bot.-London 47:755-758.
- Ridge I (1991). Plant physiology: The regulation of plant growth. Hodder and Stoughton. The Open University. pp. 7:282-333.
- Rodoni S, Muhlecker W, Anderl M, Krautler B, Moser D., Thomas H, Matile P, Hortensteiner S (1997). Chlorophyll breakdown in senescent chloroplasts (cleavage of pheophorbidea in two enzymic steps). Plant Physiol. 115:669-676.
- Sağlam-Çağ S, Cevahir G, Ünal M, Kaplan E, Çıngıl Ç, Kösesakal T (2004). The effect of Zn, Cu, Mn on senescence in excised cotyledons of *Eruca sativa* L. Fresen. Environ. Bull. 13:733-739.
- Sanchez-Bravo J, Ortuna A, Botia JM, Acosta M, Sabater F (1991). Lateral diffusion of polarly transported indoleacetic acid and its role in the growth of *Lupinus albus* L. hypocotyls. Planta 185:391-396.
- Sanders EJ, Write MA (1995). Programmed cell death in development. Int. Rev. Cytol. 163:105-173.
- Schmid M, Simpson D, Giet C (1999). Programmed cell death in castor bean endosperm is associated with the accumulation and release of a cysteine endopeptidase from ricinosomes. PNAS 96:14159-14164. Senescence and death of plant organs: nutrient recycling and developmental regulation. CR. Biol. 333:382-91.
- Shimizu-Sato S, Tanaka M, Mori H (2009). Auxin-cytokinin interactions in the control of shoot branching. Plant Mol. Biol. 69:429-435.
- Shimomura S, Inohara N, Fukui T, Futai M (1988). Different Properties of two Types of Auxin-binding Sites in Membranes from Maize Coleoptiles. Planta 175:558-566.
- Skoog F (1940). Relationships between zinc and auxin in the growth of higher plants. Am. J. Bot. 27:939-951.
- Takaki H, Kushizaki M (1970). Accumulation of free triptophan and triptamine in zinc deficient maize seedlings. Plant Cell Physiol. 11:793-804.
- Thompson JE, Froese CD, Madey E, Smith MD, Hong Y (1998). Lipid metabolism during plant senescence. Prog. Lipid. Res. 37:119-141.
- Wareing PF, Seth AK (1967). Ageing and senescence in the whole plant. In HW Woolhouse, ed, Aspects of the Biology of Ageing. Symposium of the Society for experimental Biology, Academic Press, New York 21:543-558.